

pH-driven biosynthesis of silver nanoparticle colloids from Garcinia mangostana L. (mangosteen) rind extract

UCSI-Cheras Low Carbon Innovation Hub Research Consortium

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Priority Topic Area: Responsible Production, Innovation and Industry

1 - Problem statement and aim

Two-thirds of the mangosteen fruit is still disposed as **solid waste** when it contains valuable polyphenols such as anthocyanins and flavonoids [1]. This study aimed to valorize mangosteen rind for the biosynthesis of silver nanoparticle (AgNP) colloids with pH as the driving force. This study also elucidated the yield and stability of bio-silver which are underreported in literature. The bio-silver can be potentially applied in antibacterial ultrafiltration systems for the food and beverage **industry** in future.

2 - Methodology

As illustrated in Scheme 1, the biosynthesis of AgNP colloids was performed via a facile onepot method reacting 5 mM AgNO₃ with mangosteen rind extract (9:1 v/v) upon stirring at 30 °C. 0.1 M NaOH was used to vary the pH of the medium to 8, 9, and 10, respectively.



(a) Washed rind

(d) Maceration with

10 % (v/v) ethanol

(b) Peeled exocarp

dark conditions



(e) Maceration in (f) Vacuum filtration

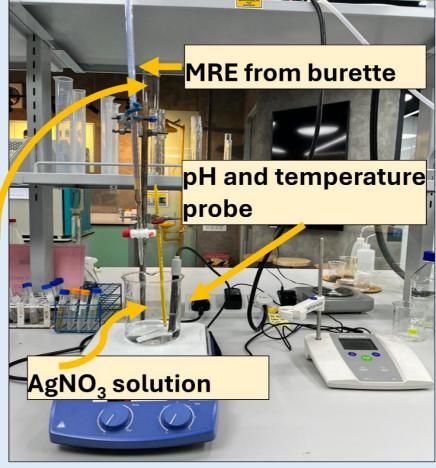
(c) Dry blending

of rind extract



(g) Mangosteen rind

extract (MRE)



(h) Bio-silver synthesis

Sample A - pure MRE (pH 3.89)

Sample B - pure NaOH

Sample C - MRE (pH 8.0)

Sample D - MRE (pH 9.0)

Sample E - MRE (pH 10.0)

UV-VIS analysis > SPR Hydrodynamic particle size/ Z-average (nm) Zeta potential (mV)

FTIR

Scheme 1. Preparation of rind extract via maceration of mangosteen exocarp with 10% (v/v) ethanol and bio-silver synthesis

3 - Results and discussion

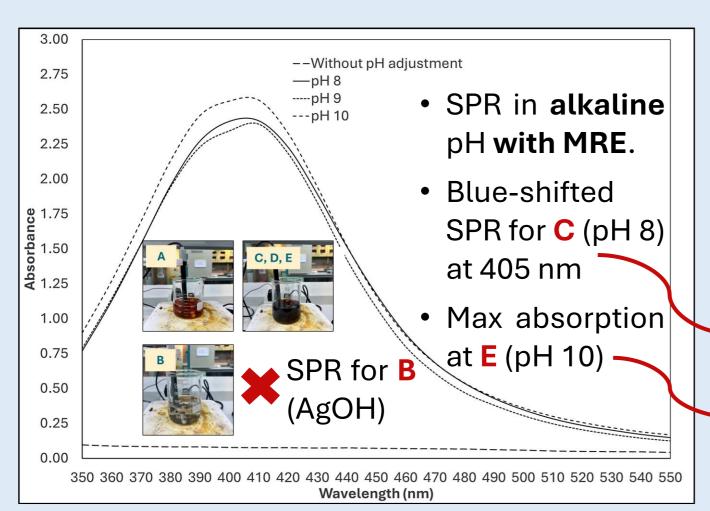


Table 1. Size and stability of MRE/AgNP colloids

	Z-average (nm)	PDI	ζ (mV)
pH 3.89 (A)	1015.50±12.50	0.44±0.07	-10.41±1.39
pH 8 (C)	128.67±0.58	0.41±0.01	-39.57±0.42
pH 9 (D)	199.07±1.20	0.50±0.03	-25.57±0.09
pH 10 (E)	200.27±2.84	0.39±0.01	-30.97±0.33

Table 2. Yield of MRE/AgNP colloids

	Yield (mg/mL)
pH 3.89 (A)	0.32±0.13
pH 8 (C)	0.47±0.14
pH 9 (D)	0.55±0.03
pH 10 (E)	0.73±0.16

- Ionic strength > steric hinderance at high pH leading to bigger sizes (agglomeration)
- **Smallest** hydrodynamic particle size and **best stability** (most negative ζ) at pH 8 (C) due to enhanced steric repulsion of bioactive compounds
- **Highest yield** at pH 10 (E) due to abundance of **-OH** groups which accelerated the reduction of Ag+ to Ag0 thus increasing AgNP nucleation [2]

Figure 1. Surface plasmon resonance (SPR)

- Metallic **Ag** bond for all samples at 409.04 (1) and 475.06 cm⁻¹ (3)
- N-O symmetry of AgNO₃ at 1057.77 cm⁻¹ (5) disappeared in Samples A, C, D, and E signifying bio-reduction by MRE
- Chelation of Ag⁺ ions with –OH (13) from carboxylic acids in mangosteen rind at **alkaline pH**
- Broadening of 957.30 (4) and 1174.02 (6) cm⁻¹ at C (pH 8) signified bond formation between Ag+ and C-N, C-O-C, C-OH moieties of flavonoids and terpenoids in MRE [3]
- Capping interactions by C-N (8) and C=O (9) groups were most pronounced at \mathbf{E} (pH 10) (increased ζ from D to E in Table 1)

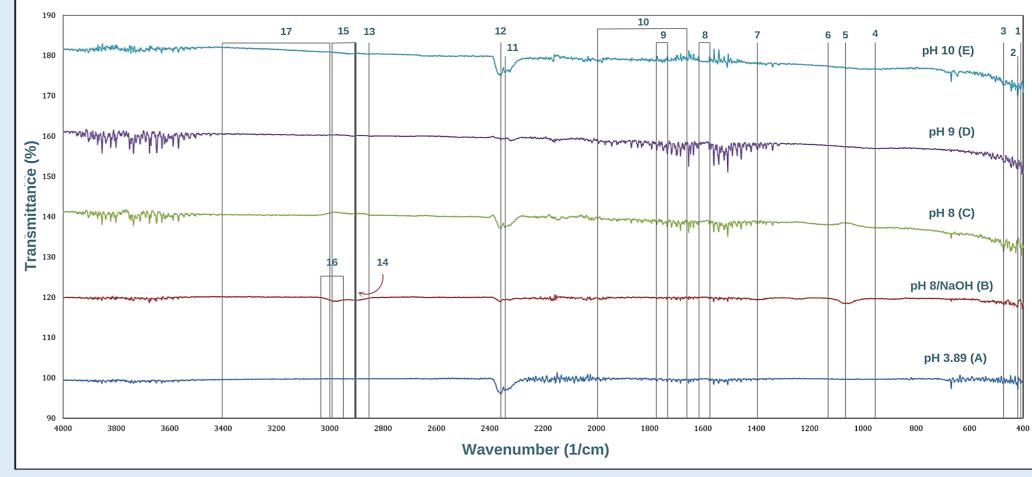


Figure 2. IR Spectra of Samples A-E

4 - Benefits to society

Repurposed mangosteen waste acts as a **sustainable bio-reductant** for the green synthesis of AgNP colloids which can serve as antibacterial agents in fabricating low-fouling ultrafiltration membranes for the food and beverage industry. Additionally, valorizing mangosteen rind lowers the environmental footprint of disposing mangosteen waste in Malaysia and Southeast Asia.

5 - Conclusion and next steps

pH 8 is recommended as the optimum pH for a future feasibility study on using bio-silver in an antibacterial ultrafiltration membrane for concentrating anthocyanins in fruit extracts.



References/Acknowledgements

[1] C. Palakawong, P. Sophanodora, P. Toivonen, and P. Delaquis, "Optimized extraction and characterization of antimicrobial phenolic compounds from mangosteen (Garcinia mangostana L.) cultivation and processing waste," J Sci Food [2] R. Zong, X. Wang, S. Shi, and Y. Zhu, "Kinetically controlled seed-mediated growth of narrow dispersed silver nanoparticles up to 120 nm: secondary nucleation, size focusing, and Ostwald ripening," Physical Chemistry Chemical Physics, vol. 16, no. 9, pp. 4236-4241, 2014. [3] V. A. Litvin and B. F. Minaev, "Spectroscopy study of silver nanoparticles fabrication using synthetic humic substances and their antimicrobial activity," Spectroschim Acta A Mol Biomol Spectrosc, vol. 108, pp. 115–122, 2013.

